

Effects of hypoxia stress on gill remodeling and apoptosis of blunt snout bream Megalobrama amblycephala "Pujiang No.2"

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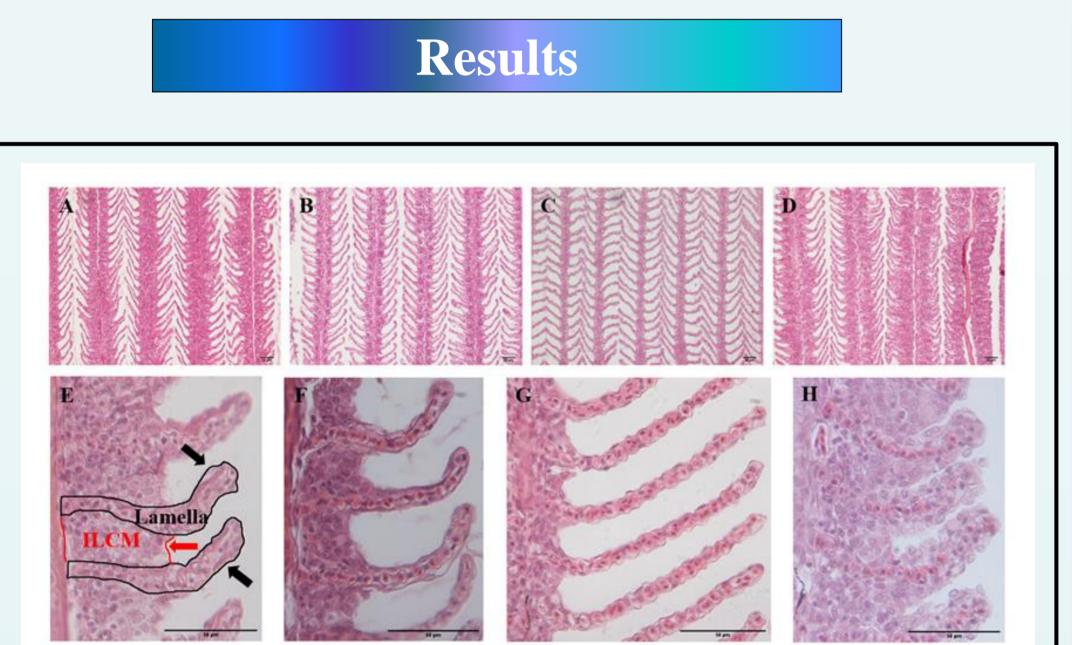


Abstract

Blunt snout bream (*Megalobrama amblycephala*) is an important species of freshwater aquaculture, but it is also a hypoxia-sensitive fish. Since 2007, the selective breeding under hypoxia was performed on a wild blunt snout bream population from Poyang Lake, China, and the new breed "Pujiang No.2" was obtained in 2020, which showed markedly improved hypoxia-tolerance. In this study, the changes of morphology and apoptosis in blunt snout bream "Pujiang No.2" gills were studied after exposed to 4, 7 days of hypoxia. Additionally, the Bcl-2 and Bad were firstly identified in blunt snout bream, and their molecular characteristics were analyzed.

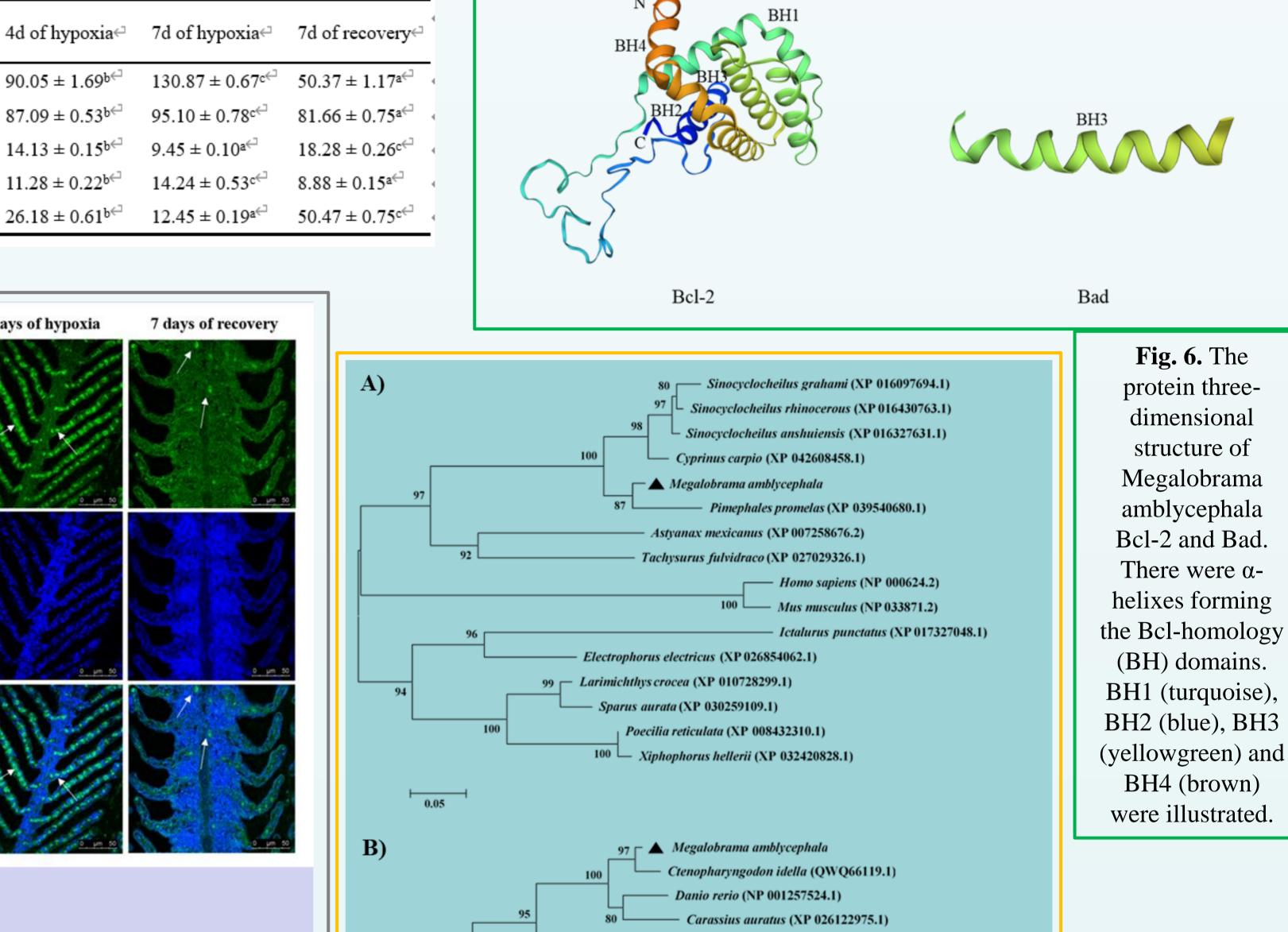
Materials and Methods	Table 2← Morphometric characteristics of blunt snout bream "Pujiang No.2" gills under hypoxia and recovery treatment.				
	Gill morphometry (25°C, DO of hypoxia was kept at 2.0 mg·L ⁻¹)	4		Megalobrama amblycephala	
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Twenty juvenile fish were exposed to hypoxia for up to 7 days at 25 °C. Five fish were taken out for sampling at 0, 4, 7 days of exposure and 7 days of recovery, respectively. Light microscopy (LM) and scanning electron microscopy (SEM) were used to observe the changes of gill morphology and structure, and morphometric analysis was carried out. TUNEL staining was used to detect the apoptosis of gills. Additionally, the Bcl-2 and Bad were firstly identified in blunt snout bream, and their molecular characteristics were analyzed. qRT-PCR was used to analyze the changes of *Bcl-2* and *Bad* mRNA expression levels in gills under hypoxia.



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Protruding lamella height (µm)⇔		← 48.12 ±	1.33ª ^{∈⊐} 90	0.05 ± 1.69 ^{b∈⊐}	130.87 ± 0.67c ^{∈∃}	50.37 ±
Protruding lamella basal length (µm)⇔		(μm)⇔ 80.28 ±	0.54ª [←] 87	7.09 ± 0.53 ^b ^{∈⊐}	95.10 ± 0.78c ^{∈∃}	81.66 ±
Protruding lamella thickness (µm)⇔		m)⇔ 18.77 ±	0.31c [←] 14	4.13 ± 0.15 ^b ^{∈⊐}	9.45 ± 0.10ª ^{∉⊐}	18.28 ±
Distance between lamellae (µm)⇔		a)⇔ 8.41 ± 0	.26ª [⇐] 11	.28 ± 0.22 ^b [←]	14.24 ± 0.53c ^{∈∃}	8.88±
ILCM height ((μm)⇔	53.79 ±	1.26c [←] 26	5.18 ± 0.61 ^b [←]	12.45 ± 0.19ª [←]	50.47 ±
(A)	Normoxia	4 days of hypoxia	n 7 days	s of hypoxia	7 days of recovery	
TUNEL				<u>0 m 10</u>		
DAPI						
Merged						
	(B) 80 (a) 60		с	□ 0 d-] □ 4 d-]		

Normoxia⇔



Pangasianodon hypophthalmus (XP 026791642.1)

Fig. 1. Light microscope (LM) micrographs of the gill lamellae for blunt snout bream "Pujiang No.2" under hypoxia and recovery treatment. Picture series starts with normoxia (A, E), 4 days of hypoxia (B, F), 7 days of hypoxia (C, G), and 7 days of recovery in normoxic water (D, H).

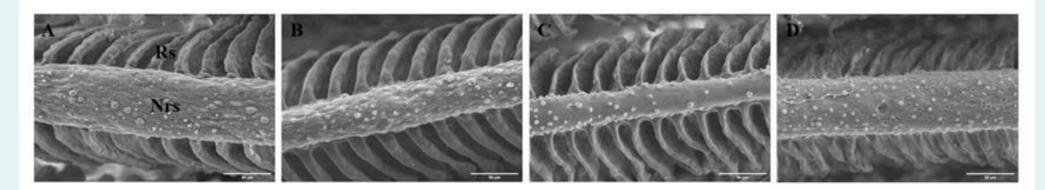
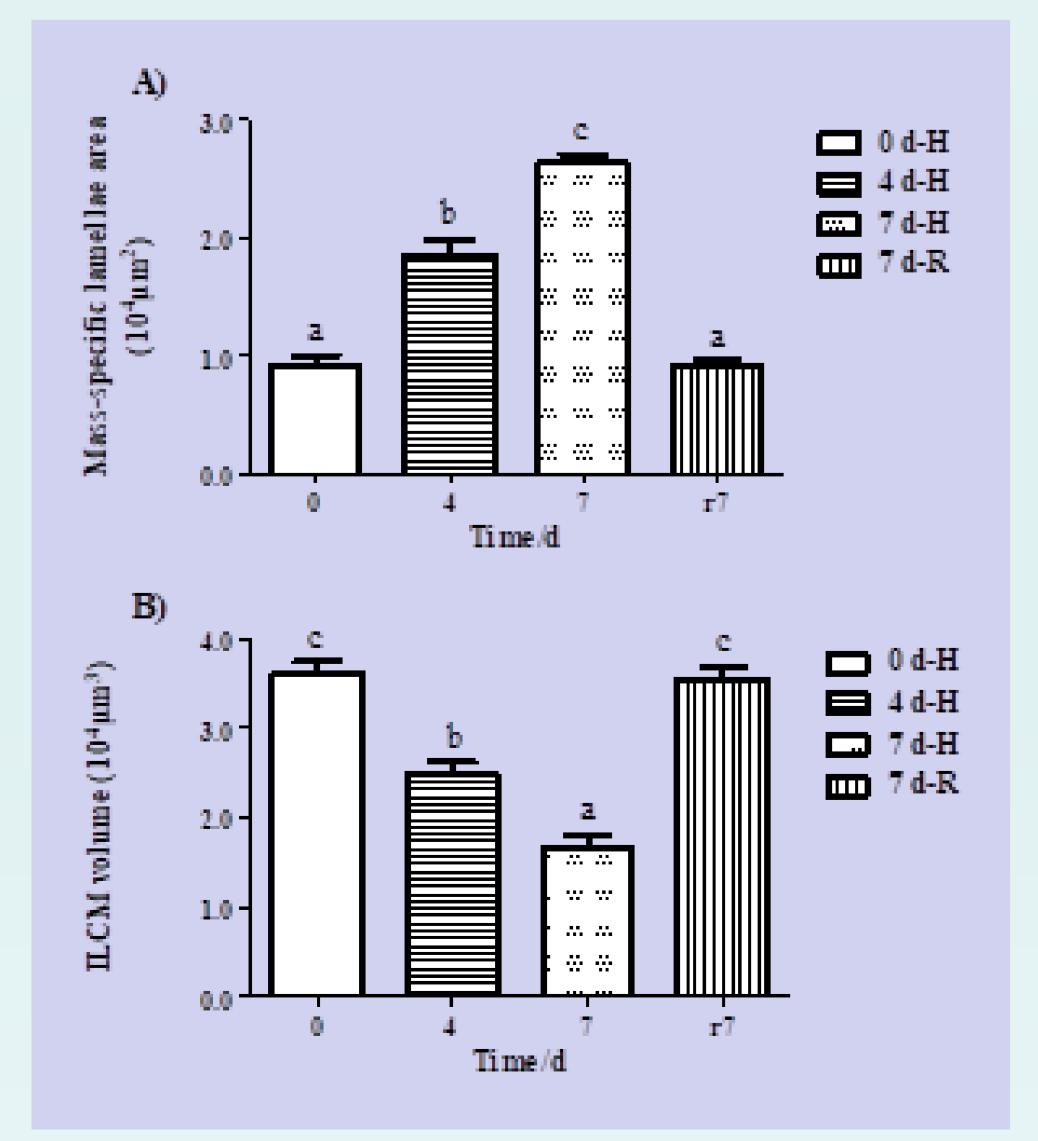


Fig. 2. Scanning electron microscopy (SEM) micrographs of the gill lamellae for blunt snout bream "Pujiang No.2" under hypoxia and reoxygenation. A-D respectively show that blunt snout bream were exposed to hypoxia for 0, 4, 7 days and returned to normoxia for 7 days.



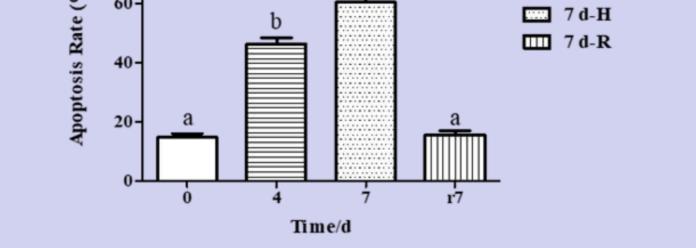
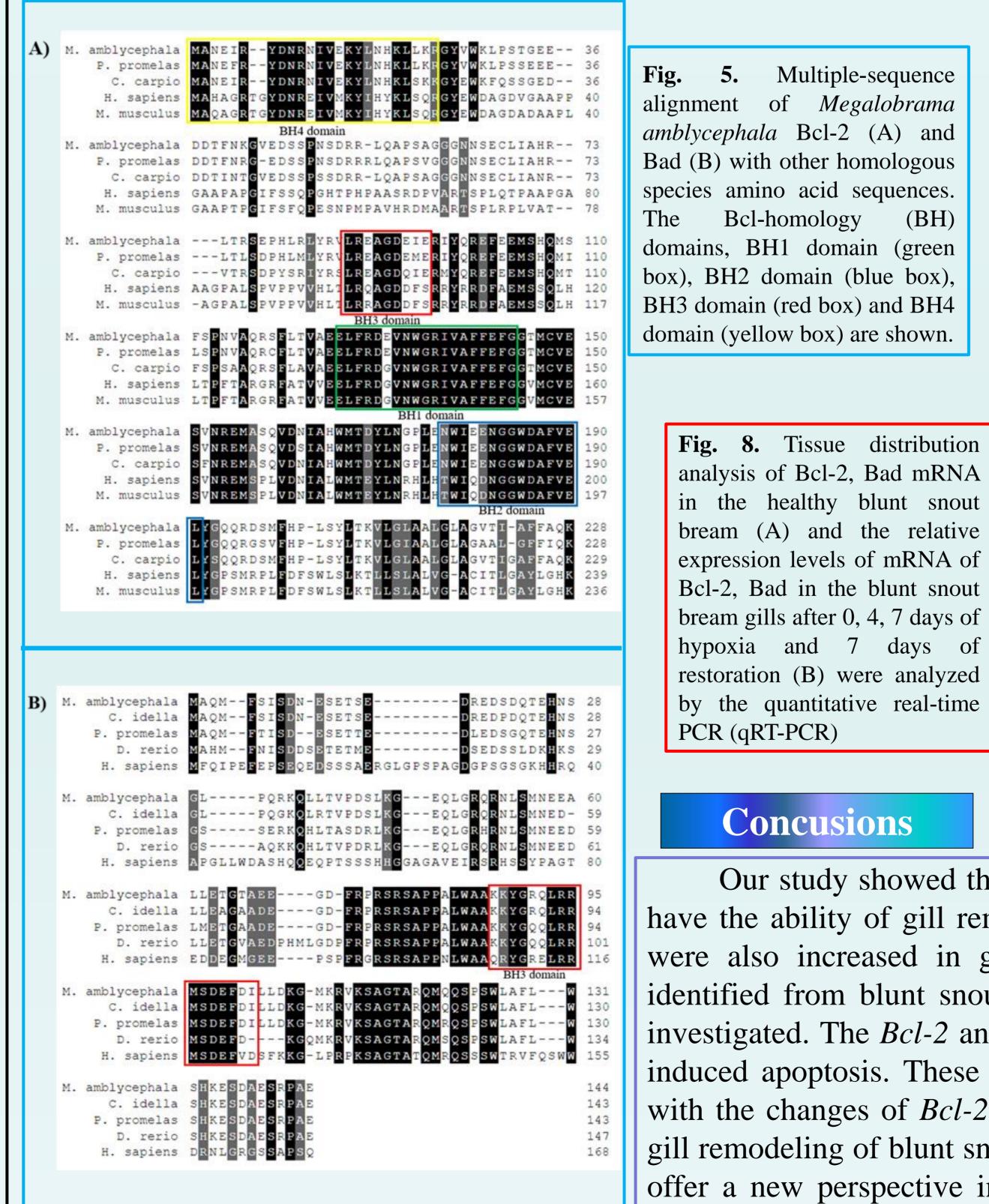
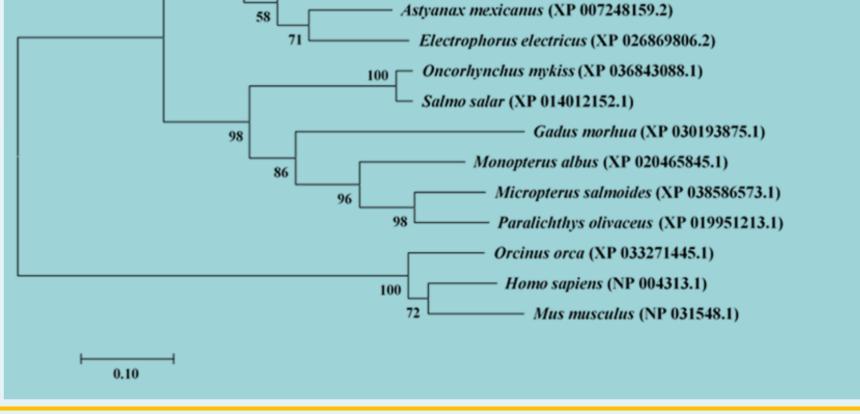
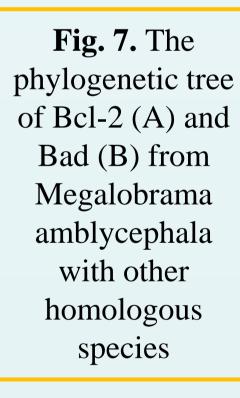


Fig. 4. Apoptosis (A) and apoptosis rate (B) of blunt snout bream "Pujiang No.2" gills under hypoxia and recovery. The apoptotic cells showed green fluorescence by TUNEL staining, all the cells showed blue fluorescence by DAPI staining.







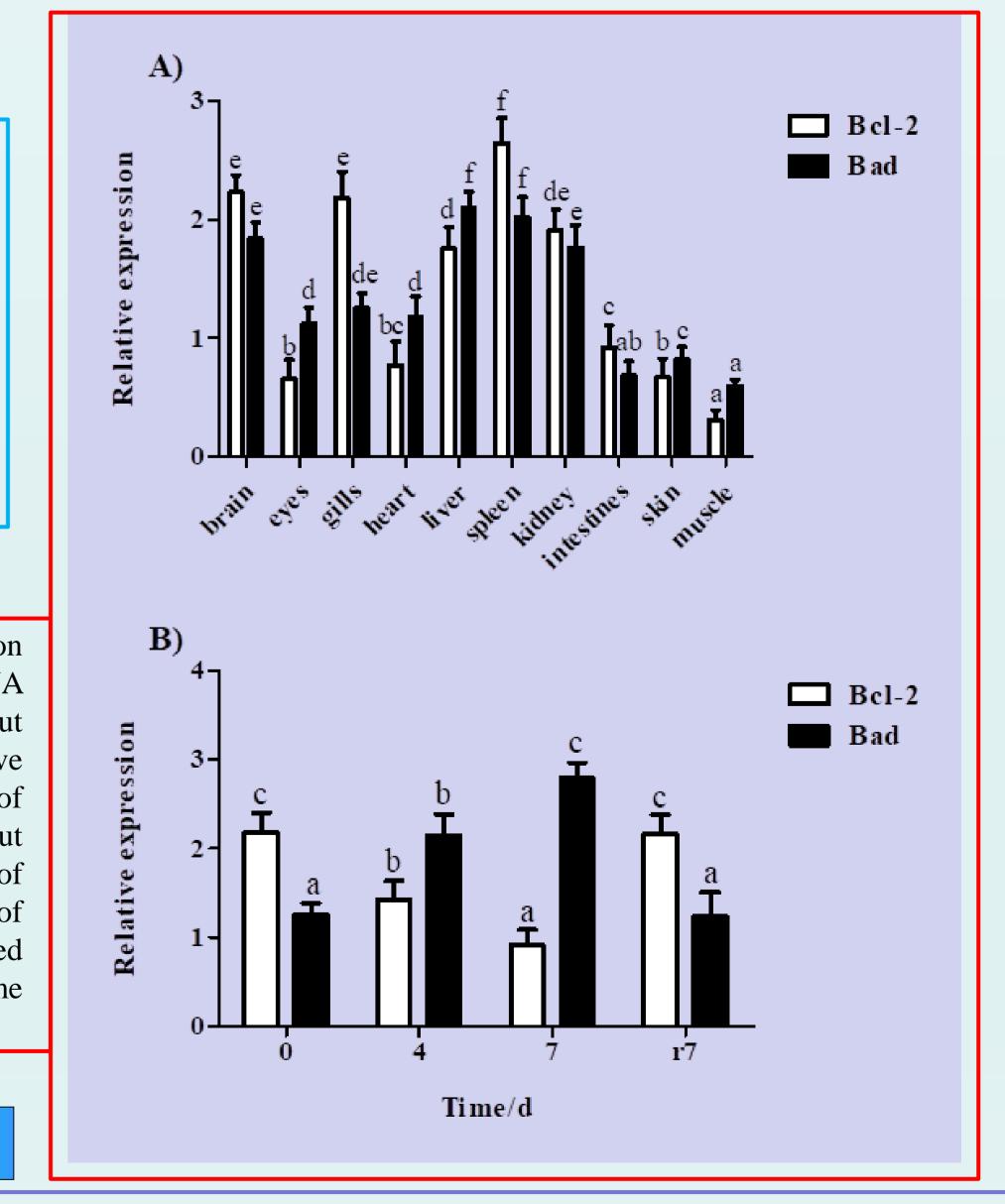


Fig. 3. Mass-specific lamellae area (A) and ILCM volume (B) change of blunt snout bream "Pujiang No.2" under hypoxia and restoration. 0 d-H: normoxia. 4 d-H: 4 days of hypoxia. 7 d-H:7 days of hypoxia. 7 d-R:7 days of recovery.

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Our study showed that the new variety "Pujiang No.2" of blunt snout bream have the ability of gill remodeling in response to hypoxia. The apoptosis signals were also increased in gills under hypoxia. The Bcl-2 and Bad were firstly identified from blunt snout bream and were found widely present in all tissues investigated. The *Bcl-2* and *Bad* may be participated in the regulation of hypoxiainduced apoptosis. These findings indicate that the increase of apoptosis signals with the changes of *Bcl-2* and *Bad* expression levels might be contributed to the gill remodeling of blunt snout bream under hypoxia stress. Collectively, our results offer a new perspective into the cellular and molecular mechanism of hypoxiainduced gill remodeling in teleost fishes.